## NOTES

## Sch 420789: A Novel Fungal Metabolite with Phospholipase D Inhibitory Activity

M. S. PUAR\*, E. BARRABEE, M. HALLADE and M. PATEL

Schering-Plough Research Institute, 2015 Galloping Hill Road, Kenilworth, New Jersey 07033-0539 USA

(Received for publication March 8, 2000)

In the course of screening for novel natural products, a fermentation culture broth (SCF0953) of an unidentified fungus showed strong activity in the phospholipase D (PLD) inhibitor assay. Phospholipase D is a lipolytic enzyme involved in the hydrolysis of phosphate bond of the phospholipid substrate generating phosphatidic acid. PLD activation is implicated in a wide range of biological activities such as inflammation, antitumor, and

antimicrobial<sup>1)</sup>. The culture sample was described as a fungal with sterile, dematiaceous mycelium with low, dry, small blastos<sup>2)</sup>. The fermentation broth was extracted with EtOAc at harvest pH (6.5). The combined extract was purified on column chromatography followed by reverse phase HPLC. The purified solid was a white powder. We

Fig. 1. The structure of **1** with relative stereochemistry.





Fig. 2. Partial structures of 1.



report herein the structure determination of this novel PLD active compound, Sch 420789 (1).

Compound 1 has the molecular formula  $C_{27}H_{36}O_6$  [*m/z* 457.2 (M+H)<sup>+</sup>] by MS, <sup>1</sup>H and <sup>13</sup>C NMR data for a total of ten degrees of unsaturation and UV (MeOH)  $\lambda_{max}$  266 nm. The <sup>1</sup>H (400 MHz, CDCl<sub>3</sub>, values in parenthesis) and <sup>13</sup>C (75.5 and 100 MHz, CDCl<sub>3</sub>) NMR spectra and their extensive analysis using APT, DEPT, <sup>1</sup>H-<sup>1</sup>H and <sup>1</sup>H-<sup>13</sup>C COSY led to the assignment of following partial structures shown in Fig. 2.

Extensive selective INEPT<sup>3)</sup> studies allowed the construction of the plain structure shown in Fig. 3. The mass spectrum (SIMS) indicated major ions at m/z 479.2 (M+Na)<sup>+</sup>, 457.2 (M+H)<sup>+</sup>, 261.1 (-196; loss of fatty acid, C<sub>12</sub>H<sub>20</sub>O<sub>2</sub>), 179 (-196 and -82; C<sub>12</sub>H<sub>20</sub>O<sub>2</sub> and C<sub>5</sub>H<sub>6</sub>O) consistent with assigned structure.

The relative stereochemistry of **1** was determined on the basis of <sup>1</sup>H-<sup>1</sup>H coupling constants and NOESY experiments. Thus, the 2'-trans, 4'-trans conjugated dienoate system of the fatty acid side chain was established on the basis of large coupling constants of  $J_{2',3'}=15.0$  Hz,  $J_{3',4'}=11.0$  Hz, and  $J_{4',5'}=15.0$  Hz. In the NOESY spectrum, the presence of a cross peak between H<sub>14</sub> and H<sub>9</sub> suggested that the methyl and the methine protons are *cis* to each other. H<sub>9</sub> was assigned  $\beta$ -axial because of its large J values with H<sub>8</sub> (J=14.0, 4.0 Hz). The lack of cross peak between  $H_{14}$  and  $H_1$  suggested *trans* relationship. The  $H_{14}$  protons also showed cross peaks to  $\beta$ -axial protons of  $H_2$  through 1,3 interactions. The protons  $H_4$  and  $H_6$  showed allylic connectivity *via* W-coupling in the COSY spectrum and a cross peak observed in the NOESY spectrum between  $H_4$ and  $H_6$  provided justification for the hydroxy group being  $\beta$ -axial at  $C_4$ . On the basis of above data the structure of **1** was assigned.

Compound 1 showed *in vitro* inhibitory activity in the PMA- and *f*MLP-stimulated phospholipase (PLD) assay<sup>4</sup>). The IC<sub>50</sub> values of 1 were ~20 and ~8  $\mu$ M, respectively. Compound 1 lacked selectivity.

## References

- BILLAH, M. M.: Phospholipase D and cell signalling. Curr. Opin. Immunol. 5: 114~123, 1993
- The microorganism was supplied by Dr. B. KATZ of MYCOsearch Lab (MYCO 0929)
- For a recent application of this technique towards structure elucidation see; PETTIT, G. R., SINGH, S. B., GOSWAMI, A., NIEMAN, R. A. Tetrahedron 44: 3349, 1988
- PAI, J.-K.; E. A. FRANK, C. BLOOD & M. CHU: Novel ketoepoxides block phospholipase D activation and Tumor cell invasion. Anti-Cancer Drug Design 9: 363~372, 1994